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Technical overview – Automated high-throughput HMW DNA extraction for PacBio long-read sequencing using Nanobind HT kits

Sequel II and IIe systems ICS v11.0 Revio system ICS v13.0+ SMRT Link v13.0+

Automated high-throughput high-molecular weight (HMW) DNA extraction for PacBio long-read sequencing using Nanobind HT kits

Technical Overview

- 1. Nanobind HT HMW DNA extraction workflow overview
- 2. Nanobind HT HMW DNA extraction workflow details
- 3. Nanobind HT HMW DNA extraction example performance data
- 4. Technical documentation & applications support resources

Nanobind high-throughput (HT) HMW DNA extraction for PacBio long-read sequencing: Getting started



Nanobind DNA extraction technology overview

Nanobind technology enables extraction of high-molecular weight DNA from common samples as well as more challenging samples such as animal tissue, insects and plants





Extracted HMW DNA visibly bound to a Nanobind disk

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Nanobind HT HMW DNA extraction workflow overview

Nanobind HT kits enable automated high-throughput HMW DNA extraction for PacBio HiFi sequencing

Nanobind HT kits are designed for human/animal blood, mammalian cells, and bacteria, and are compatible with KingFisher robotic platforms



Automated solutions built on Nanobind technology

Nanobind HT kits use magnetic disk processing to automate lysis, binding, washing and elution steps and are compatible with instruments from Hamilton and Thermo Fisher

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- Hamilton NIMBUS Presto is a walkaway solution with automated plate filling
- Thermo Fisher KingFisher instruments are semi-automated with manual plate filling and limited user interaction during the run



Available Nanobind HT kits

Nanobind HT CBB kit (102-762-700; 96 rxn)

- For up to 200 μL human/mammalian blood, nonmammalian animal blood¹, cultured cells, and bacteria
- Expected HMW DNA yield: 3–15 µg for blood and cultured mammalian cells and 2–10 µg for bacteria

Nanobind HT 1 mL blood kit (102-762-800; 96 rxn)

- For 1 mL human blood
- Expected HMW DNA yield: 3–70 µg

How Nanobind HMW DNA extraction kits work with KingFisher systems

Kingfisher sample purification systems employ inverse magnetic particle processing (MPP) technology to enable automated transfer and processing of magnetic Nanobind disks in a microplate format.

Kingfisher instruments



- Automate sample extraction by moving magnetic particles or objects (not liquids)
- Inverse MPP uses magnetic rods covered with a disposable tip comb and PCR or deepwell plates
- Function without any liquid dispensing or aspiration parts or devices
- Enable quicker reaction times and a more efficient wash process

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How Nanobind HMW DNA extraction kits work with KingFisher systems (cont.)

Standard manual processing with Nanobind disks

Liquid is moved into and out of a tube using a pipette while the magnetic Nanobind disk remains immobilized inside the tube





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Automated inverse magnetic particle processing (MPP) with Nanobind disks

Rather than moving liquids, **magnetic Nanobind disks are moved** from plate to plate using magnetic rods covered with a disposable plastic tip comb



Schematic illustration of KingFisher rotary platform modified *after* Ficarro et al., Anal Chem. 2009 June 1; 81(11): 4566–4575. doi:10.1021/ac9004452.

Nanobind HT HMW DNA extraction workflow overview



* Refer to appropriate vendor websites to download user guides and other documentation for specific automation systems.

PacBio scripts support high-throughput HMW DNA extraction using Nanobind HT kits with semi-automated and fully-automated workflows



Available Nanobind HT HMW DNA extraction protocols

Select the appropriate Nanobind HT Procedure & checklist to use based on the type of automation system, sample type, desired sample throughput and available input sample amount

Automation	Dropoduro ⁹ obseklist	Sample ture	Sample	Max #	Nanobind HT kit (96-rxn)	
system	Procedure & checklist	Sample type	input	run	Kit	Part no.
KingFisher Duo	Extracting HMW DNA using the Nanobind HT CBB kit for mammalian cultured cells on KingFisher Duo Prime system [102-996-200]	Mammalian cells	1x10 ⁶ cells ¹	12	Nanobind HT CBB	102-762-700
	Extracting HMW DNA using the Nanobind HT CBB kit for 200 μL human whole blood on KingFisher Duo Prime system [102-995-800]	Human whole blood	200 μL	12	kit	102-702-700
÷	Extracting HMW DNA using Nanobind HT 1 mL blood kit for human whole blood on KingFisher Duo Prime system [102-995-400]	Human whole blood	1 mL	6	Nanobind HT 1 mL whole blood kit	102-762-800
KingFisher Flex	Extracting HMW DNA using the Nanobind HT CBB kit for mammalian cultured cells on KingFisher Flex system [<u>102-996-300]</u>	Mammalian cells	1x10 ⁶ cells ¹	96	Nanobind HT CBB	102 762 700
	Extracting HMW DNA using the Nanobind HT CBB kit for 200 μL human whole blood on KingFisher Flex system [102-995-900]	Human whole blood	200 μL	96	kit	102-102-100
	Extracting HMW DNA using Nanobind HT 1 mL blood kit for human whole blood on KingFisher Flex system [102-995-500]	Human whole blood	1 mL	24	Nanobind HT 1 mL whole blood kit	102-762-800
KingFisher Apex	Extracting HMW DNA using the Nanobind HT CBB kit for mammalian cultured cells on KingFisher Apex system [102-996-100]	Mammalian cells	1x10 ⁶ cells ¹	96		102-762-700
	Extracting HMW DNA using the Nanobind HT CBB kit for 200 μL human whole blood on KingFisher Apex system [102-995-700]	Human whole blood	200 μL	96		
	Extracting HMW DNA using the Nanobind HT CBB kit for bacteria on the KingFisher Apex system [103-377- 600]	Cultured bacteria	0.5 mL (OD600 = 1) ²	96	Nanobind HT CBB kit	
	Extracting HMW DNA using the Nanobind HT CBB kit for non-human mammalian blood (NHMB) on the KingFisher Apex system [103-397-300]	Non-human mammalian blood	200 μL	96		
	Extracting HMW DNA using the Nanobind HT CBB kit for nucleated red blood cells (nRBCs) on the KingFisher Apex system [103-377-800]	Non-mammalian blood (nucleated RBCs)	5 – 20 μL	96		
	Extracting HMW DNA using Nanobind HT 1 mL blood kit for human whole blood on KingFisher Apex system [102-995-300]	Human whole blood	1 mL	24	Nanobind HT 1 mL whole blood kit	102-762-800

PacBie ¹ Cultured mammalian cell samples are typically resuspended in 50 µL of 1X PBS for Nanobind HT processing.

² For many bacteria, 1.0 OD600 will typically correspond to ~1 x 10⁹ cells per mL, but the actual cell density range is large across all species.

Available Nanobind HT HMW DNA extraction protocols (cont.)

Select the appropriate Nanobind HT Procedure & checklist based on the type of automation system, sample type, desired sample throughput and available input sample amount

Automation system	Dropoduro 9 obsoklist	Somela tura	Input	Max #	Nanobind HT kit (96-rxn)	
		Sample type	amount	run	Kit	Part no.
Hamilton NIMBUS	Extracting HMW DNA using the Nanobind HT CBB kit for mammalian cultured cells on Hamilton NIMBUS Presto system [102-996-400]	Mammalian cells	1x10 ⁶ cells ¹	96		
Presto	Extracting HMW DNA using the Nanobind HT CBB kit for 200 μL human whole blood on Hamilton NIMBUS Presto system [102-996-000]	Human whole blood	200 μL	96		
	Extracting HMW DNA using the Nanobind HT CBB kit for bacteria on Hamilton NIMBUS Presto system [103- 397-400]	Cultured bacteria	0.5 mL (OD600 = 1) ²	96	Nanobind HT CBB kit	102-762-700
	Extracting HMW DNA using the Nanobind HT CBB kit for non-human mammalian blood (NHMB) on the Hamilton NIMBUS Presto system [<u>103-377-700]</u>	Non-human mammalian blood	200 μL	96		
	Extracting HMW DNA using the Nanobind HT CBB kit for nucleated red blood cells (nRBCs) on the Hamilton NIMBUS Presto system [103-397-500]	Non-mammalian blood (nucleated RBCs)	5 – 20 μL	96		
	Extracting HMW DNA using the Nanobind HT 1 mL blood kit for human whole blood on Hamilton NIMBUS Presto system [102-995-600]	Human whole blood	1 mL	24	Nanobind HT 1 mL whole blood kit	102-762-800

See PacBio's Documentation website for the most up-to-date list of Nanobind HT HMW DNA sample preparation protocols



¹ Cultured mammalian cell samples are typically resuspended in 50 µL of 1X PBS for Nanobind HT processing.

² For many bacteria, 1.0 OD600 will typically correspond to \sim 1 x 10⁹ cells per mL, but the actual cell density range is large across all species.

Mammalian versus non-mammalian animal blood

For Nanobind HT HMW DNA extraction procedures, mammalian blood samples typically require higher input volumes than non-mammalian blood samples

Mammalian blood

- Mammalian red blood cells (RBCs) typically do not contain nuclei and thus cannot be used for DNA extraction
- For mammalian blood samples, typically extract DNA from white blood cells (WBCs)
- We recommend using 200 µl of whole blood (fresh or frozen) for DNA extraction from mammalian blood samples using Nanobind kits

Non-mammalian blood

Most non-mammalian vertebrate animals like birds, fish, and reptiles have nucleated red blood cells that can be used for DNA extraction

→ Can reduce non-mammalian blood input sample volume ~10-fold to 80-fold compared to mammalian blood samples

We recommend using **2.5 – 20 µl of whole blood** (fresh or frozen) for DNA extraction from non-mammalian blood samples using Nanobind kits



Extraction from 200 µL of mammalian blood

Extraction from 2.5–20 µL of non-mammalian blood

Supported automation platforms for high-throughput HMW DNA extraction using Nanobind HT kits

Nanobind HT HMW DNA extraction kits are optimized for use with Thermo Fisher Scientific KingFisher sample purification systems and Hamilton Microlab NIMBUS automated liquid handlers



	Thermo Fisher Scientific KingFisher Duo Prime	Thermo Fisher Scientific KingFisher Flex	Thermo Fisher Scientific KingFisher Apex	Hamilton Microlab NIMBUS Presto
Instrument design	 Compact benchtop sample purification system 	 Benchtop sample purification system 	 Benchtop sample purification system with touchscreen 	 Hamilton liquid handler with integrated KingFisher Presto sample purification system
Samples per run	• 6 or 12	• 24 to 96	• 24 to 96	• 24 to 96
Nanobind HT workflow automation level	 Semi-automated Manual plate filling + requires limited user interaction after run start 	 Semi-automated Manual plate filling + requires limited user interaction after run start 	 Semi-automated Manual plate filling + requires limited user interaction after run start 	 Fully automated Automated plate filling + fully walk- away after run start
Processing time	<2 hrs	~2 hrs	~2 hrs	~2.5 hrs
Automation run time	1 – 1.8 hrs	1 – 1.8 hrs	1 – 1.8 hrs	~2 hrs
Hands-on time	~15 min	~20 – 30 min	~20 – 30 min	~25 min
Supported sample types	 Human blood (200 µL and 1 mL) Cultured mammalian cells 	 Human blood (200 µL and 1 mL) Cultured Mammalian cells 	 Human blood (200 µL and 1 mL) Animal blood (mammalian and non-mammalian) Cultured Mammalian cells Cultured bacteria 	 Human blood (200 µL and 1 mL) Animal blood (mammalian and non-mammalian) Cultured Mammalian cells Cultured bacteria

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Nanobind HT HMW DNA extraction workflow details

Nanobind HT Procedure & checklist documentation overview

Nanobind HT Procedures & checklists support high-throughput HMW DNA extraction workflows on KingFisher and Hamilton automation systems

Nanobind HT Procedure & checklist protocols describe procedures for automated HMW DNA extraction from human/animal blood, mammalian cells, and bacteria using a Kingfisher Duo/Flex/Apex system or Hamilton NIMBUS Presto system

Procedure & checklist contents

- 1. Equipment and reagent list
- 2. Sample input requirements
- 3. Automated HMW (50 kb 300+ Mb) DNA extraction protocol
- 4. QC procedure

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- 5. Expected QC results
- 6. Troubleshooting FAQ

Nanobind HT Procedures & checklists have been validated using human/animal whole blood samples, bacteria and mammalian cells



Extracting HMW DNA using the Nanobind[®] HT CBB kit for mammalian cultured cells on the KingFisher Apex system Procedure & checklist **PacBi**

This procedure describes the workflow for high-throughput automated extraction of HMW (50–300 kb) DNA from cultured mammalian cells using the Thermo Fisher KingFisher Apex robotic instrument. This procedure requires the Nanobind HT CBB kit (102-762-700) and is recommended for HiT sequencing.

The Nanobind HT CBB kit has enough reagents for 96 extractions to be run in one of the following formats: 1 run x 96 samples, 2 runs x 48 samples, or 4 runs x 24 samples. We do not recommend running fewer than 24 samples per run as the kit is designed to accommodate dead volumes for a maximum of 4 runs (4 runs x 24 samples).

Required equipment and materials

Equipment/reagent	Manufacturer (part number)
Nanobind HT CBB kit	PacBio® (102-762-700)
KingFisher Apex System	Thermo Fisher Scientific (5400930, includes Apex 96 deep well magnet head)
KingFisher Apex 96 deep-well magnet head	Thermo Fisher Scientific (24079930)
KingFisher Apex 96 deep-well heating block	Thermo Fisher Scientific (24075920)
KingFisher 96 deep-well plates, barcoded	Thermo Fisher Scientific (95040450B)
KingFisher 96 deep-well tip combs, barcoded	Thermo Fisher Scientific (97002534B)
Ethanol (96–100%)	Any major lab supplier (MLS)
Isopropanol (100%)	Any MLS
UV/Vis	Thermo Fisher Scientific NanoDrop 2000
Fluorescent DNA Quantification	Thermo Qubit 3.0, dsDNA BR and RNA BR Assay Kits

Before you begin

Prior to starting

Buffer CW1 and CW2 are supplied as concentrates. This kit uses CW1 with a 60% final ethanol concentration. This kit uses CW2 with a 60% final ethanol concentration. Before using, add the appropriate amount of ethanol (96-100%) to Buffer CW1 and Buffer CW2 as indicated on the bottles.

See PacBio sample preparation Documentation

See PacBio's Documentation website for the most up-to-date list of Nanobind HT HMW DNA sample preparation protocols.

Equipment & materials required for Nanobind HT HMW DNA extraction

For each type of sample purification system, use the correct matching magnet heads, heating blocks, plates and other consumables as listed in the appropriate Nanobind HT procedure



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* Thermo Fisher Scientific PN 24078841 = KingFisher Presto 24 DW head (24078840) + one 24 DW heating block (24075440)

Sample input requirements for Nanobind HT HMW DNA extraction from human whole blood and mammalian cells

	Human whole blood	Mammalian cells			
Nanobind HT kit	Nanobind HT CBB kit (96 Rx) (102-762-700) Nanobind HT 1 mL whole blood kit (96 Rx) (102-762-800)	Nanobind HT CBB kit (96 Rx) (102-762-700)			
Storage conditions	 Use Potassium EDTA (K₂EDTA) anticoagulant Sodium heparin (NaHep) and citrate (NaCit) also performed well in limited testing Store at 4°C for ≤2 days to prevent sample degradation 	• N/A			
	 Cells: White blood cell (WBC) count should be ≥4 × 10⁶ cells/mL* to achieve ≥3 ug DNA yield for input into SMRTbell library prep 	 Use 1x10⁶ diploid human cells* or equivalent Determine cell counts using a hemocytometer or cell counter 			
Sample input amount	 Blood: Use either 200 μL or 1 mL of input human whole blood 200 μL: DNA yield is 1 – 12 μg depending on donor WBC conc. 1 mL: DNA yield is 3 – 70 μg depending on donor WBC conc. 	 For non-diploid/non-human cells: Scale cell input to contain 5–25 µg of DNA Warning: >25 µg inputs can cause Nanobind disks to be "dropped" in Lysis/Binding solution and/or cause well-to-well contamination 			
Sample freezing / thawing conditions	 Freeze blood samples as quickly as possible after being drawn Aliquot blood samples to avoid repeated freeze-thaws Thaw blood samples in water bath/dry block heater at 37 °C (15 min Mix by inverting tube >15 times immediately prior to use Note: Improperly thawed and mixed samples may result in inconsistent DNA yield and purity 	 Cell pellets should be frozen dry with as much liquid removed as possible. No cryoprotectant needed 			
	No systematic difference observed in DNA QC or sequencing results between fresh and frozen samples				

Before you begin

End-to-end overall Nanobind HT workflow

Prepare sample, storage & reagent plates

Select appropriate automation script to run

Load prepared plates and other consumables onto work deck

Run selected Nanobind HT DNA extraction protocol

Store extracted DNA samples and perform QC

Semi-automated workflow (KingFisher Duo/Flex/Apex)





Fully-automated workflow (Hamilton NIMBUS Presto)



Prior to beginning Nanobind HT protocol

- Ensure that the proper automation script has been installed (see <u>Nanobind HT kit Guide & overview</u> "Programs" section)
 - To obtain KingFisher automation scripts, contact PacBio Technical Support at support@pacb.com
 - To obtain Hamilton NIMBUS Presto automation scripts, contact <u>Hamilton Technical Support</u> at <u>roboticservice@hamiltoncompany.com</u>
- Ensure that instrument is set up with the correct magnet head and heating block.

Before you begin (cont.)

End-to-end overall Nanobind HT workflow

Prepare sample, storage & reagent plates

Semi-automated workflow (KingFisher Duo/Flex/Apex)





Fully-automated workflow (Hamilton NIMBUS Presto)



Select appropriate automation script to run

Load prepared plates and other consumables onto work deck

Run selected Nanobind HT DNA extraction protocol

Store extracted DNA samples and perform QC

Prior to beginning Nanobind HT protocol

For Hamilton NIMBUS Presto only:

- Prepare a *.xls sample worklist
- Column 1 = "Sample_ID"

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- List one sample ID for each sample to be processed
- Column 2 = "Sample_Position"
 - List the corresponding well location for each sample

Sample worklist enables NIMBUS liquid handler to **automatically** prepare lysis reactions on instrument work deck using a loaded sample plate

	Α	В	С	D
1	Sample_ID	Sample_Position		
2	Sample1	A1		
3	Sample2	B1		
4	Sample3	C1		
5	Sample4	D1		
6	Sample5	A2		
7	Sample6	B2		
8	Sample7	C2		
9	Sample8	D2		
10	Sample9	A3		
11	Sample10	B3		
12	Sample11	C3		
13	Sample12	D3		
14	Sample13	A4		
15	Sample14	B4		

Sample, storage & reagent plate preparations

NIMBUS Presto workflow requires upfront preparation of **fewer plates** compared to Duo/Flex/Apex workflows

End-to-end overall Nanobind HT workflow Prepare sample, storage & reagent plates Load prepared plates and other consumables onto work deck Run selected Nanobind HT DNA

Store extracted DNA samples

and perform QC

Semi-automated workflow (KingFisher Duo/Flex/Apex)

Fully-automated workflow (Hamilton NIMBUS Presto)

Manually prepare deep-well plates with required quantities of Nanobind reagents & consumables

Example list of 7 DW plates to be prepared for Kingfisher Apex.

Plate	Plate name	Reagent	Notes
1	Lysis / binding	Sample / lysis / binding solution	Prepare only after all other plates have been prepared (see below)
2	Nanobind storage	1 Nanobind disk per well*	200 μL / cells \rightarrow 3 mm disk 1 mL \rightarrow 5 mm disk
3	Wash 1	Buffer CW1	200 μL sample / cells \rightarrow 700 μL 1 mL sample \rightarrow 2 mL
4	Wash 2	Buffer CW2	200 μL sample / cells \rightarrow 700 μL 1 mL sample \rightarrow 2 mL
5	Wash 3	Buffer CW3	200 μL sample / cells \rightarrow 700 μL 1 mL sample \rightarrow 2 mL
6	Elution	Buffer EB	200 μL sample / cells \rightarrow 100 μL 1 mL sample \rightarrow 200 μL
7	Tip comb storage	For 24- or 96-tip comb	Insert tip comb into a DW plate for loading onto instrument

Plate 1 (lysis / binding plate) preparation procedure :

- Add Proteinase K
- 2. Add whole blood sample
- 3. Add Buffer BL3
- 4. Add RNase A

Note: Sample and reagents MUST be added to each well in the order described in the Procedure

Example list of 3 DW plates to be prepared for NIMBUS Presto.

Plate	Plate name	Reagent	Notes
1	Sample plate	Samples for analysis	Prepare only after all other plates have been prepared
2	Nanobind storage	1 Nanobind disk per well*	200 μL sample / cells $\ \rightarrow$ 3 mm disk 1 mL sample \rightarrow 5 mm disk
3	Tip comb storage	For 24- or 96-tip comb	Insert tip comb into a DW plate for loading onto instrument

Plate 1 (sample plate) preparation procedure:

1. Add whole blood sample to appropriate well positions as specified in *.xls sample worklist.



1	A	В	С	D
1	Sample_ID	Sample_Position		
2	Sample1	A1		
3	Sample2	81		
4	Sample3	C1		
5	Sample4	D1		
6	Sample5	A2		
7	Sample6	B2		
8	Sample7	C2		
9	Sample8	D2		
10	Sample9	A3		
11	Sample10	B3		
12	Sample11	C3		
13	Sample12	D3		
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Pace * Nanobind disks do not need to be perfectly centered, but please ensure they are at the bottom of the well and not stuck to the side walls.

Automation script selection



Work deck preparation



Automation script execution

End-to-end overall Nanobind HT workflow

Prepare sample, storage & reagent plates

Select appropriate automation script to run

Load prepared plates and other consumables onto work deck

Run selected Nanobind HT DNA extraction protocol

Store extracted DNA samples and perform QC

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Semi-automated workflow (KingFisher Duo/Flex/Apex)

The run will start <u>automatically</u> when final plate is loaded and **Next** button is pressed.



After run starts, screen shows time until user action required, remaining run time, and current protocol step



User action required: When prompted, remove Lysis/Binding plate (Plate 1) from instrument and manually add IPA to each well. After adding IPA, return plate to instrument and resume program.



Note: Add isopropanol (IPA) gently against the **side** of the well into the Lysis / Binding solution.*

Fully-automated workflow (Hamilton NIMBUS Presto)

Close front cover of instrument to protect against environmental contamination and click **Ok** to start run



NIMBUS Presto laptop screen will request final verification that correct magnet head is installed prior to starting run



No user action required until run completes

* Adding IPA directly to the Lysis / Binding solution may affect extraction purity.

Sample storage and QC

End-to-end overall Nanobind HT workflow

Prepare sample, storage & reagent plates

Select appropriate automation script to run

Load prepared plates and other consumables onto work deck

Run selected Nanobind HT DNA extraction protocol

Store extracted DNA samples and perform QC

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Semi-automated workflow (KingFisher Duo/Flex/Apex)

Fully-automated workflow (Hamilton NIMBUS Presto)

When run ends, remove each plate from instrument as prompted.











Manually transfer eluates from Elution Plate to a new storage plate or storage tubes



Elution plate New storage plate tube

KingFisher Duo/Flex/Apex automation scripts are designed to leave Nanobind disks in elution plate after program completes

NIMBUS Presto automation scripts are designed to place Nanobind disks back into the disk storage plate, so the elution plate should not contain any disks inside

Pipette mix extracted DNA sample 10 times with a standard P200 pipette to homogenize and disrupt any unsolubilized "jellies" that may be present. Take care to disrupt any regions that feel more viscous than other regions.



Note: Limited pipette mixing will not noticeably reduce DNA size or sequencing read lengths but is important for accurate quantitation and consistent sequencing performance

Let eluate rest overnight at room temp. to allow DNA to solubilize. Following overnight rest, pipette mix 10 times with a standard P200 pipette. Visible "jellies" should disperse after resting.

Recommended QC procedure for HMW DNA samples extracted using Nanobind HT kits

Use recommended tools to perform DNA QC evaluation of high-molecular weight DNA



Qubit fluorometer

DNA concentration QC

Use Qubit dsDNA broad range (BR) assay kit for DNA concentration QC

• We recommend using a Qubit fluorometer (Thermo Fisher Scientific) with the dsDNA BR assay kit since it provides more reliable concentration measurements of (unsheared) HMW DNA

Optional: Determine RNA concentration using the Qubit RNA BR assay kit (Thermo Fisher Scientific)

• We recommend taking a single measurement to obtain an approximate RNA concentration reading



Nanodrop

DNA purity QC

Perform UV/VIS assay measurements with a Nanodrop to determine total nucleic acid conc. as well as purity (A260/A280, A260/230)

- Note: Inconsistent Nanodrop concentration or spectrophotometric readings may be obtained if the DNA is very heterogeneous or contains large amounts of unsolubilized "jellies"
- For information on how to disrupt particularly viscous regions, refer to the kit Guide & overview "Heterogeneity and viscosity" section or the "Troubleshooting FAQ" section in the Procedures & checklists



Femto Pulse system

DNA sizing QC

Use a Femto Pulse system with Genomic DNA 165 kb Kit (Agilent Technologies) for HMW genomic DNA sizing QC

- We recommend diluting samples to 250 pg/µL (use wide-bore pipette for making dilutions and gentle mixing)
- · Avoid mixing with a standard pipette! This will shear dilute solutions of DNA.

Expected QC results for HMW DNA samples extracted using Nanobind HT kits

Example ranges typically observed for DNA concentration, DNA purity and DNA sizing QC metrics



Qubit fluorometer

DNA concentration QC

- 200 µL human whole blood typically yields ~3 12 µg of DNA depending on donor white blood cell (WBC) count
- 1 mL of human whole blood typically yields ~3 70 µg depending on donor WBC count
- 1x10⁶ GM24385 cells typically yields ~4 10 µg
- 1x10⁶ MCF-7 cells typically yields ~12 18 μg



Nanodrop

DNA purity QC

- 260/280 ratios should consistently be 1.8 2.0
- 260/230 ratio can vary from 1.3 2.2
- Samples with UV purities within expected ranges should sequence well
- UV purities outside of these ranges may indicate abnormalities in extraction process



Femto Pulse system

DNA sizing QC

 Mode of extracted human cell and whole blood DNA measured on Femto Pulse system (Agilent Technologies) is typically 100 kb+

Refer to Nanobind HT Procedure & checklist documentation for guidance on troubleshooting specific types of DNA extraction quality issues

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Nanobind HT HMW DNA extraction example performance data

Nanobind HT kit example performance

Example Nanobind HT DNA extraction QC and HiFi sequencing performance results¹



Example Femto Pulse DNA sizing QC analysis results for HMW DNA samples extracted with Nanobind HT kit using KingFisher Apex automation platform.





Example HiFi read length distribution for HG001 cell sample extracted with Nanobind HT kit using Hamilton NIMBUS Presto system.

Nanobind HT kit example performance (cont.)

Example Nanobind HT DNA extraction QC and HiFi sequencing performance results¹

Sample	Input material	DNA yield	DNA mode size	HiFi mean read length	HiFi yield
Whole human blood	200 µL	4.5 μg	135 kb	18,109 bp	92 Gb
Whole human blood	1 mL	28.4 μg	114 kb	15,132 bp	102 Gb
Mammalian cultured cell (HG001)	1 x 10 ⁶ cells	5.9 μg	86 kb	17,393 bp	105 Gb
Bovine blood	200 μL	15 μg	50 kb	15,988 bp	106 Gb
Chicken blood (nucleated red blood cell)	2.5 μL	11.5 μg	137 kb	16,384 bp	98 Gb
Bacteria G-, <i>E. Coli</i>	5 x 10 ⁸ cells	5 µg	106 kb	9427 bp 2	67 Ch2
Bacteria G+, L. monocytogenes	5 x 10 ⁸ cells	8 µg	122 kb	0437 bp-	07 GD-

² Microbial gDNA samples were sheared to 8 kb mean fragment size according to recommendations, whereas other samples shown in table were sheared to 15–18 kb. Following SMRTbell library preparation, microbial samples were multiplexed for sequencing on a single Revio SMRT Cell.

SMRTbell prep kit 3.0 (SPK 3.0) WGS SMRTbell libraries constructed with Nanobind HT-extracted DNA show excellent HiFi sequencing performance



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Technical documentation & applications support resources

Technical resources for Nanobind HT HMW DNA extraction

Nanobind HT Procedure & checklist documentation

Automation system	Procedure & checklist	Sample type
KingFisher Duo	Extracting HMW DNA using the Nanobind HT CBB kit for mammalian cultured cells on KingFisher Duo Prime system [102-996-200]	Mammalian cells
	Extracting HMW DNA using the Nanobind HT CBB kit for 200 μL human whole blood on KingFisher Duo Prime system [102-995-800]	Human whole blood
	Extracting HMW DNA using Nanobind HT 1 mL blood kit for human whole blood on KingFisher Duo Prime system [102-995-400]	Human whole blood
KingFisher Flex	Extracting HMW DNA using the Nanobind HT CBB kit for mammalian cultured cells on KingFisher Flex system [102-996-300]	Mammalian cells
	Extracting HMW DNA using the Nanobind HT CBB kit for 200 μ L human whole blood on KingFisher Flex system [102-995-900]	Human whole blood
	Extracting HMW DNA using Nanobind HT 1 mL blood kit for human whole blood on KingFisher Flex system [102-995-500]	Human whole blood
KingFisher Apex	Extracting HMW DNA using the Nanobind HT CBB kit for mammalian cultured cells on KingFisher Apex system [102-996-100]	Mammalian cells
	Extracting HMW DNA using the Nanobind HT CBB kit for 200 μ L human whole blood on KingFisher Apex system [102-995-700]	Human whole blood
	Extracting HMW DNA using the Nanobind HT CBB kit for bacteria on the KingFisher Apex system [103-377-600]	Cultured bacteria
	Extracting HMW DNA using the Nanobind HT CBB kit for non-human mammalian blood (NHMB) on the KingFisher Apex system [103-397-300]	Non-human mammalian blood
	Extracting HMW DNA using the Nanobind HT CBB kit for nucleated red blood cells (nRBCs) on the KingFisher Apex system [103-377-800]	Non-mammalian blood (nucleated RBCs)
	Extracting HMW DNA using Nanobind HT 1 mL blood kit for human whole blood on KingFisher Apex system [102-995-300]	Human whole blood

Technical resources for Nanobind HT HMW DNA extraction (cont.)

Nanobind HT Procedure & checklist documentation

Automation system	Procedure & checklist	Sample type
Hamilton NIMBUS Presto	Extracting HMW DNA using the Nanobind HT CBB kit for mammalian cultured cells on Hamilton NIMBUS Presto system [102-996-400]	Mammalian cells
	Extracting HMW DNA using the Nanobind HT CBB kit for 200 µL human whole blood on Hamilton NIMBUS Presto system [102-996-000]	Human whole blood
	Extracting HMW DNA using the Nanobind HT CBB kit for bacteria on Hamilton NIMBUS Presto system [103-397-400]	Cultured bacteria
	Extracting HMW DNA using the Nanobind HT CBB kit for non-human mammalian blood (NHMB) on the Hamilton NIMBUS Presto system [103-377-700]	Non-human mammalian blood
	Extracting HMW DNA using the Nanobind HT CBB kit for nucleated red blood cells (nRBCs) on the Hamilton NIMBUS Presto system [103-397-500]	Non-mammalian blood (nucleated RBCs)
	Extracting HMW DNA using the Nanobind HT 1 mL blood kit for human whole blood on Hamilton NIMBUS Presto system [102-995-600]	Human whole blood

Nanobind HT HMW DNA sample preparation literature

- Brochure Nanobind HT HMW DNA extraction (102-326-565)
- Guide & overview Nanobind HT kits for automated HMW DNA extraction (<u>103-028-100</u>)
- Technical note High-throughput DNA extraction (<u>102-326-611</u>)
- Technical overview Automated high-throughput HMW DNA extraction for PacBio long-read sequencing using Nanobind HT kits (<u>103-401-</u><u>700</u>)

Technical resources for Nanobind HT HMW DNA extraction (cont.)

Third-party automation instrumentation literature

- Hamilton Technical Note Automated extraction of High Molecular Weight (HMW) DNA with PacBio Nanobind technology on the Hamilton NIMBUS Presto Assay Ready Workstation (<u>AN-2205-05</u>)
- Hamilton Technical Note Automated Isolation of High Molecular Weight (HMW) DNA from Human Blood Samples with PacBio Nanobind Technology on the Hamilton NIMBUS Presto - Next Level Preparation of Extracts for Long-Read Sequencing (<u>AN-2212-03</u>)

Posters

• PacBio poster (2022) – High-throughput workflow for human whole genome sequencing using PacBio HiFi [Link]

Nanobind HT consumable product literature

- Nanobind HT 1 mL blood kit (PN <u>102-762-800</u>)
 - Product insert: Nanobind HT 1 mL blood kit 4C
 - Product insert: Nanobind HT 1 mL blood kit RT
- Nanobind HT CBB kit (PN <u>102-762-700</u>)
 - Product insert Nanobind HT CBB kit 4C
 - Product insert Nanobind HT CBB kit RT
- PacBio sample prep consumables website [Link]

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