HIGHLY ACCURATE HIFI READS FOR GENE EDITING VALIDATION

With highly accurate long reads (HiFi reads) from the Sequel[®] II and Sequel IIe system, powered by Single Molecule, Real-Time (SMRT[®]) sequencing technology, you can comprehensively validate gene editing techniques including adeno-associated virus (AAV) and CRISPR-Cas9 approaches.

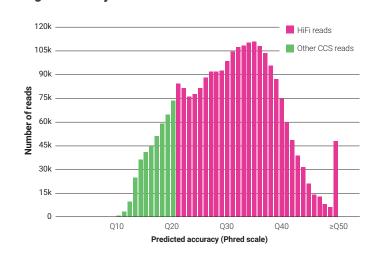
Sequence with confidence

High accuracy

- Discover novel potential capsid constructs
- Conduct vector quality control experiments
- Detect and accurately measure the efficiency of both on- and off-target effects
- Assess the safety of resulting constructs

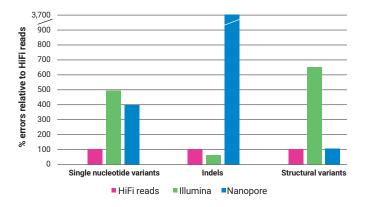
The advantages of HiFi reads for gene editing validation

- Long read lengths up to 25 kb to span complete genes or regions of interest
- High accuracy of 99.9% (Q30) to provide Sangerquality, base-level resolution
- Comprehensively detect all variant types to ensure both base-level modifications and structural rearrangements are captured
- Uniform coverage to detect variants in repetitive and extreme GC-regions in an unbiased manner



Data from a 15 kb size-selected human library using the SMRTbell® express template prep kit 2.0 on a Sequel IIe system (2.0 chemistry, Sequel IIe system software v10, 30-hour movie).

Comprehensive variant detection



Variant calling performance against *Genome in a Bottle* benchmarks for PacBio HiFi reads (35-fold, Sequel II system, 2.0 chemistry); Illumina (35-fold, NovaSeq); Oxford Nanopore (60-fold, PromethION R9.4.1).



editing outcomes
 Develop and

Characterize CRISPR-Cas9

Develop and assess AAV- _ based therapeutic approaches

- 11			_		
- 88					
- 11					
	-	_	_	_	
	_	_	_	_	
	_		_	_	
	-	_		_	
	_		_		
			_		
		_	_	_	
	_	_	_	_	
	_	_	_	_	
		_			

Use amplification

or amplificationfree approaches

Workflow

With easy-to-use, high throughput sequencing, you can get the accuracy you need at an affordable cost



Sample prep

Use single-stranded DNA (ssAAV), selfcomplementary DNA (scDNA)^{1,2,3} or DNA amplicons as starting material.



Library prep

Use standardized and automatable workflows for a range of target types with multiplexing options for up to 96 targets.



SMRT sequencing

Use 10–20 hr movies to generate 1–3 million HiFi reads up to 20 kb in length per SMRT® Cell 8M on Sequel II or IIe systems.

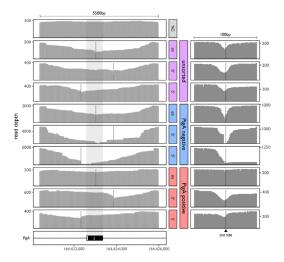


Data analysis

Use SMRT[®] Link, a webbased end-to-end workflow manager to demultiplex, analyze, and visualize your sequencing data.

Sequence beyond your target

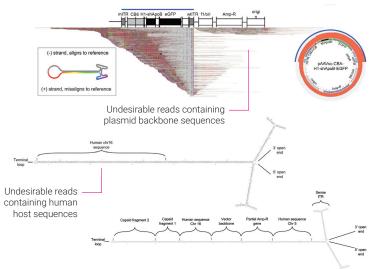
Understanding the extent of CRISPR-Cas9 editing requires long read lengths and high accuracy to capture both on- and off-target effects to fully evaluate editing outcomes.^{4,5,6,7}



Analysis of the PigA locus edited with the CRISPR-Cas9 method using selected gRNAs. SMRT sequencing of a 5.5 kb amplicon around gRNA cut sites (vertical lines) enabled detection of large-scale deletions and structural changes missed by other methods.⁶

Assess gene transfer vectors

Performing quality control of AAV vectors with highly accurate long reads enables complete sequencing of viral particles to ensure correct packaging.^{4,5,8}



SMRT sequencing provided a complete sequence of viral particles for heterogeneous vector populations to the pCis-plasmid reference. This allowed for unambiguous detection of plasmid and host gDNA that was encapsidated.[®]

KEY REFERENCES

- 1. Guerin, K. et al. (2020) A novel next-generation sequencing and analysis platform to assess the identity of recombinant adeno-associated viral preparations from viral DNA extracts. *Human Gene Therapy*. 31(11-12):664-678.
- Lecompte, E. et al. (2015) Advanced characterization of DNA molecules in rAAV vector preparations by single-stranded virus next-generation sequencing. Molecular Therapy Nucleic Acids. 4:E260.
- 3. Gao, G. and Sena-Esteves, M. (2020) Introducing genes into mammalian cells: viral vectors. Cold Spring Harbor Spring Protocol. (8):095513
- 4. Tran, N. T. et al. (2020) **AAV-genome population sequencing of vectors packaging CRISPR components reveals design-influenced heterogeneity.** *Molecular Therapy*. Methods & Clinical Development, 18, 639–651.
- 5. Larrea, A. and Tai, P.W.L. (2019) Highly accurate SMRT sequencing for gene editing applications. PacBio webinar.
- Kosicki, M. et al. (2018) Repair of double-strand breaks induced by CRISPR-Cas9 leads to large deletions and complex rearrangements. Nature Biotechnology, 36, 765–771.
 Höijer, I. et al. (2020) Amplification-free long-read sequencing reveals unforeseen CRISPR-Cas9 off-target activity. Genome Biology. 21, 290.
- Tai, P. W. L. et al. (2018) Adeno-associated virus genome population sequencing achieves full vector genome resolution and reveals human-vector chimeras. *Molecular Therapy: Methods & Clinical Development*. Vol 9, 130-141.

Learn about highly accurate HiFi reads for gene editing validation: pacb.com/target

Information in this document is subject to change without notice. PacBio assumes no responsibility for any errors or omissions in this document. Certain notices, terms, conditions and/or use restrictions may pertain to your use of PacBio products and/or third party products. Refer to the applicable PacBio terms and conditions of sale and to the applicable license terms at http://www.pacb.com/legal-and-trademarks/terms-and-conditions-of-sale/. PacBio, the PacBio logo, SMRT, SMRTbell, and Sequel are trademarks of PacBio. All other trademarks are the sole property of their respective owners.

PacBi

© 2022 PacBio. All rights reserved. Research use only. Not for use in diagnostic procedures. 102-193-695 REV01 18MAR2022